REGIOSELECTIVE SYNTHESES OF ETHER-LINKED PORPHYRIN DIMERS AND TRIMERS RELATED TO PHOTOFRIN-II@

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Abstract Using isomerically pure 4-(1-hydroxyethyl)deuteroporphyrin-IX dimethyl ester 10, 2-(1 hyrhoxyethyl)deuteroporphyrin-IX dimethyl ester **11,2-acetyl-4-(l-hydmxyethyl)deuteroporphyrin-IX** dimethyl ester 12 and 4-acetyl-2-(l-hydroxyethyl)deuteroporphyrin-IX dimethyl ester 13 as starting materials, a series of regicchemically pure ether-linked porphyrin dimers 25-41 and trimers 42-47 related to an anticancer drug known as Photofrin-II@ were synthesized. Proton NMR nuclear Overhauser enhancement experiments and variable temperature proton NMR spectroscopy were used to characterize the isomers. Some of these compounds were tested for their in vivo photosensitizing ability vis-a-vis Photofrin-II@ and the preliminary results are briefly described.

One of the most promising new modalities being explored for use in the control and treatment of tumors is "Photodynamic Therapy" (PDT).¹ This treatment is based on the interaction of a photosensitizer retained in malignant tissue with photons of visible light, resulting in the formation of singlet oxygen, the putative lethal agent.² At present, Photofrin-II@, a purified form of hematoporphyrin derivative is the only drug which is being evaluated in Phase-III clinical trials in the United States, specifically for treatment of obstructive endobronchial tumors and superficial bladder tumors.

The commercial method for preparation of Photofrin-II® is depicted in Scheme $1³$ In brief, hematoporphyrin-IX 1 is first treated with a sulfuric acid/acetic acid mixture to produce mainly hematopotphytin-IX diacetate 2 and the corresponding mono-acetates 3 and 4. Treatment of this mixture with aqueous sodium hydroxide (O.lM) causes hydrolysis and elimination to produce compounds S-P. Porphyrins 2-9 subsequently couple to provide a gross mixture termed "hematoporphyrin derivative" (Hpd). Hpd contains monomers such as hematoporphyrin-IX **1,2-** and **4-** (1-hydroxyethyl) deuteroporphyrin-IX 7,8 and protoporphyrin-IX 9, as well as higher oligomers. The aqueous solution thus obtained is separated by size exclusion gel chromatography into two fractions. The higher molecular weight fraction contains the material which is active in vivo (the so-called "Photofrin-II®")⁴ and the lower molecular weight fraction contains mainly the monomers, which are either biologically inactive or else promote adverse reactions in the skin.⁵ Kessel and co-workers have reported an alternate procedure for Hpd preparation via pure hematoporphyrin diacetate 2 by reacting hematoporphyrin-IX **1** with acetic anhydride/pyridine and then treating the product with dilute aqueous sodium hydroxide, $6\,\,\text{which ultimately resulted in the same complex mixture.}$ As can be. seen fmm the HPLC chromatogram (Figure l), Photofrin-II@ is a complex mixture in all respects. The **peaks** with retention times 5.0, 15.0, 16.5 and 25.0 min have been characterized as belonging to porphyrin **1,7,8,** and 9. The oligomeric fraction with retention times between 19 and 30 min is a complex mixture and it is now accepted that this fraction contains dimers linked with ether, $7-9$ ester, $10-12$ and/or carbon-carbon linkages.^{13,14} The recent data obtained from fast atom bombardment (PAB) mass spectroscopy indicate that the oligomeric components of Photoftin-II@ contain up to nine porphyrin units and the commericial batches of Photofrin-II@ prepared under the same controlled conditions have the same oligomeric distribution.15

10 $R^2 = H$ **;** $R^4 = CH(OH)Me$ 11 R^2 = CH(OH)Me; R^4 = H $12 R² = COMe: R⁴ = CH(OH)Me$ 13 **R2 = CH(OH)Me; R4 = COMe** 14 R^2 = COMe; R^4 = CH=CH₂ $15 R² = CH = CH₂; R⁴ = COMe$ $16R^2 = R^4 = COMe$ 17 **R2 = CH(OH)Me; R4 = CH=CH2** $18 R² = CH = CH₂; R⁴ = CH(OH)Me$ **19** R^2 **= COMe;** R^4 **= CH(Br)Me** $20 R^2 = CH(Br)Me; R^4 = COMe$ $21 R^2 = R^4 = CH(Br)Me$ $22 R²$ = CH(OCH₂OCH₂CH₂SiMe₃)Me; $23 R² = COMe;$

The chemical nature of the tumor localizing fraction of Photofrin-II@ has been a subject of discussion for quite some time. Various research groups have made attempts to separate individual components of the oligomeric material in Photofrin-II@ by using chromatography, including reversed-phase HPLC, but without much success.¹⁶ We recently reported results on a chromatographically well-separated fraction of the tumor localizing component (Figure 1, peak A),¹⁷ obtained by using a combination of gel filtration chromatography and semi-preparative high performance liquid chromatography. On the basis of mass spectrometric analysis and behavior of the fraction towards base hydrolysis and lithium aluminum hydride reduction, it was deemed to contain ether-linked porphyrin dimers along with a small amount of hematoporphyrin trimer and various dehydration (vinyl) products. The HPLC results suggest that in the oligomeric mixture of Photofrin-II@ the retention times of most of the dimers and trimers are very close and they are virtually impossible to separate. Thus, due to the complex nature of Photofrin-II@ and the difficulty in separating the individual components, efforts have been made by various groups to synthesize the possible components of this tumor localizing fraction.

Our own work to elucidate the structure of Photofrin-II@ started a R_1^4 = COMe few years ago; we have previously reported the syntheses of dimers and $\frac{1}{2}$ **R4** = CH(OCH₂CH₂CH₂SiMe₃)Me trimers with ester, ether, and carbon-carbon linkages. Dimers with ester 24 R² = CH(OCH₂CCH₂CH₂SiMe₃)Me linkages were biologically inactive, while dimers and trimers with ether **R4 = CH(OH)Me** linkages forged between the 2- and 4- positions of hematoporphyrin-IX showed significant anti-cancer activity. In our initial synthetic studies, we

started with a mixture of 2-acetyl-4-(l-hydroxyethyl)- and 4-acetyl-2-(1-hydroxyethyl)-deuteroporphyrin-IX dimethyl ester 12 and 13, respectively, which in turn were prepared as a mixture by partial reduction of 2,4 diacetyldeuteroporphyrin-IX dimethyl ester 16. This resulted in isolation of mixtures of regio- and stereo-isomeric products.⁸ Ward et al. have also reported the synthesis of similar mixtures.¹⁸

In our studies with dimers linked by carbon-carbon linkages, 19 we observed that the dimer prepared from 4- $(1$ hydroxyethyl)-deuteroporphyrin-IX dimethyl ester IO was extremely active as an anticancer agent, while the dimer obtained from 2-(1-hydroxyethyl)-deuteroporphyrin-1X dimethyl ester 11 showed little photosensitizing ability. These results prompted us to synthesize regiochemically pure dimers and trimers linked with ether linkages. We anticipate that the biological studies with pure photosensitizers will enable us to know more about structure/activity relationships among the numerous different components of Photofrin-II@.

Synthesis of regiochemically pure dimers and trimers requires the ready availability of large quantities of isomerically pure mono-acetyl-mono-(l-hydroxyethyl)-deuteroporphyrin-IX dimethyl ester isomers 12 and 13. We recently reported²⁰ an efficient synthesis and characterization of these isomers in a single step from hematoporphyrin-IX dimethyl ester by using tetrapropyl ammoniumperruthenate (TPAP) and 4-methylmorpholine-N-oxide. In our preliminary communication we briefly reported the utility of porphyrins 12 and 13 for the synthesis of porphyrin dimers and trimers. Here, we wish to present the detailed synthesis of porphyrin dimers 25-40 and trimers 41-46 with various substituents at the peripheral positions, and their characterization by proton NMR, nuclear Overhauser enhancement (NOE) experiments, fast atom bombardment mass spectrometry (FAB). high resolution mass spectrometry (HRMS), and by classical chemical degradation approaches.

31 $R^1 = R^2 = H$ $32 R^1 = R^2 = COMe$ 3 3 $R^1 = R^2 = CH(OH)Me$ 3 4 R^1 = COMe; R^2 = CH=CH₂ 35 R^1 = CH(OH)Me; R^2 = CH=CH₂

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HN

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Me

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HN

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CO₂Me

CO₂Me

fluoride trihydrate under very mild conditions.²¹ We used this strategy and dimer 30 was obtained in 32% yield starting from porphyrin 24. However, problems were encountered in cleavage of the silyl groups with fluoride ion at room temperature, and at higher temperature the desired product was obtained in poor yield along with a large number of decomposition products. In another approach, porphyrin 12 was treated with methanesulfonyl chloride (at -7O'C). The resulting mesylate was treated with lithium bromide and the (1-bromoethyl)porphyrin 19 was immediately condensed with porphyrin 12 and the diacetyl dimer 26 was obtained in 25% yield. Reduction of the acetyl groups in 26 with sodium borohydride afforded the hematoporphyrin dimer 27 in excellent yield. The acetyl dimer 26 was also prepared by converting 2-acetyl-4-(1 -hydroxyethyl)-deuteroporphyrin-IX dimethyl ester 12 into the corresponding (lbromoethyl) analogue by reaction with HBr gas in dichloromethane. The solvent was evaporated under high vacuum

 $MeO₂C$

MeO₂C

and the (I-bromoethyl) compound was immediately condensed with porphyrin 12 to give the desired dimer 26 in 50% yield. Thus, by using the appropriate starting porphyrins, a series of dimers 25-41 was synthesized. For the preparation of porphyrin trimer 42, hematoporphyrin-IX dimethyl ester was first converted into its (1-bromoethyl) analogue with HBr gas, and then condensed with two equiv of 2-acetyl-4-(1-hydroxyethyl)-deuteroporphyrin-IX dimethyl ester 12. The acetyl groups were then reduced with sodium borohydride and the hematoporphyrin trimer 43 was obtained in 75% yield. By following similar chemistry and using the appropriate starting porphyrins, a series of trimers 44-47 was synthesized in 30 to 35% yields.

Diacetyl dimers 26, 32 and 36, as individual regioisomers, were prepared as a mixture of three stereoisomers $(d, l \text{ and } meso)$ and dimers containing one acetyl and one vinyl group (28, 34, 38, 40) are individually a mixture of four possible stereoisomers. Proton NMR assignments of the resonances for various groups in diacetyl- and monoacetyl-monovinyldimers were easier to make than for the corresponding (1-hydroxyethyl)-derivatives due to addition of two more (in case of dimers 27, 33 and 37) or one more (29,35, 39,41) chiral centers. As shown in Figure 2, in the NMR spectrum of the symmetrical diacetyl

dimer 26. eight meso protons were observed at 9.08, 9.58, 9.92 and 10.38 ppm, each integrating for two protons. The broad peak observed at 9.58 ppm was assigned to two meso (β, β') protons adjacent to the ether bridge [CH(CH3)-0-CH(CH3)]; this was further confirmed by NOE studies in which irradiation of the ether bridge CH3 protons (d at δ 2.58) resulted in a NOE to the meso protons at β and β' . The CH(CH3)-O-CH(CH3) protons were observed as a single quartet at 6.61 ppm.

Unlike dimers 26 and 32, the unsymmetrical dimer 36 featured eight meso protons in the range 10.38 to 9.10 ppm. Six of these meso protons were observed as sharp singlets and two as broad peaks. Like the symmetrical diacetyl dimers 26 and 32, the broad peaks in dimer 36 were assigned to the β, α' meso protons. The CH(CH3)-O-CH(CH3) were observed as separate quartets at 6.65 and 6.60

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ppm, eachintegrating for one proton. In the diacetyl trimer 42, sixteen stereoisomers are theoretically possible; twelve meso protons were observed in the range 10.60 to 9.38 ppm (Figure 2). The CH(CH₃)-O-CH(CH₃) protons were ,observed as a multiplet from 6.80 to 5.92 ppm. All four possible regioisomers of mono-acetyl-mono-vinyl dimers 2%, 34,38,40 were prepared by following an approach similar to described previously for the diacetyl dimer. The NMR spectra of dimers 28 and 40 were similar, but were different from dimers 34 and 38. The NMR spectrum of dimer 28 shows eight meso protons (six sharp singlets and two broad peaks) in the meso region; the α -vinyl proton was observed as a multiplet at 8.00 ppm and the β -vinyl protons were observed as two doublets at 6.14 and 6.35 ppm, each integrating for one proton. Two CH protons of the bridging CH(CH3)-O-CH(CH3) moiety were assigned as a multiplet at 6.62 ppm. The meso and vinyl resonances for dimers 34 and 38 display a different pattern, and we did not assign the resonances observed at 5.26 and 5.5 1 ppm (both are doublets); this work is in progress rnd will be reported later.

Figure 2: Proton NMR spectra (low field region only, at 300 MHz) in CDCl₃ of A, dimer 36; B, dimer 26; C, trimer, 42

Classical chemical degradation approaches were also used to confirm the structures of all dimers and trimers. For example, treating dimers 25, 26 or 27 with aqueous sulfuric acid produced the related monomers such as protoporphyrin-IX 9 and the corresponding mono-acetyl-monovinyl-porphyrin 14, due to cleavage of the ether linkage and subsequent dehydration of the (1-hydroxyethyl) analogue. Diacetyl trimers 42 and 45 produced the corresponding monomers which were characterized by proton NMR spectroscopy. Hematoporphyrin trimers 43 and

 $25R^1 = R^2 = COMe$ $26 R^1 = R^2 = CH(OH)Me$ $27 R^1 = COMe$; $R^2 = CH = CH_2$

46 and divinyl trimers 44 and 47, upon similar treatment (after diazomethane treatment) produced protoporphyrin-IX dimethyl ester as the sole product. Acid treatment of these dimers afforded protoporphyrin-IX 9 as the sole product. The structures of all these dimers and trimers were also confirmed by use of FAB mass spectrometry and HRMS.

In recent years there has been much discussion about the stability of Photofrin-II@ components. In our hands, porphyrin dimers with ester linkages were found to be unstable and were readily cleaved by leaving a solution at room temperature even for a short time. In our preliminary communication we showed, by variable temperature NMR spectroscopy, that ether linked dimers and trimers are stable at high temperature, but were cleaved easily even under slightly acidic conditions (pH, 6.2). From this study it is quite possible that the ether linked oligomer of Photofrin-II@ might be cleaving in the tumor (at pH 6.4) at body temperature (37° C). Further model work as well as in vivo NMR studies with these components are in progress.

All the synthetic dimers and trimers are being tested for in vivo photosensitizing activity, vis-a-vis Photofrin-II@. The tumor system (subcutaneously implanted SMT-f tumors in DBAL? mice) and the biological testing pmcedure were followed as described by Dougherty et al.²² Normal tissue response was tested by exposing the mouse foot to light in a manner identical with the tumor response test. These photosensitizers were tested, either as methyl esters or as the corresponding carboxylic , and identical results were obtained in either case. In brief, hematoporphyrin dimers with ether linkages (27, 33, and 37) were found to be less active than Photofrin-II®. Replacement of one of the (1hydroxyethyl) groups with a vinyl group (as in dimer 29) was found to introduce moderate activity. When both heat $(43-44^{\circ}C)$ and light were used, dimer 29 was found to be extremely active, even at doses lower than used with Photofrin-II[®]. Under similar conditions, hematoporphyrin dimers 27, 33, and 37 did not show any significant increase in their photosensitizing activity. Dimers 25 and 31, in which the (1-hydroxyethyl) groups have been replaced by a hydrogen, were found to be almost as active as Photofrin-II@. Amongst the trimers, so far only hematoporphyrin trimer 43 and divinyl trimer 44 have been tested. Preliminary results show that divinyl trimer 44 is more active than trimer 43, which is much more active than the hematoporphyrin dimers 27, 33, and 37.

Our preliminary data indicate that hydrophilicity/hydrophobicity are one of the factors which play an important role in localization of photosensitizers in tumors. For example in hematoporphyrin dimers 27 , 33, and 37, two $(1$ hydroxyethyl) groups make them more hydrophilic than the mono-(1-hydroxyethyl)-mono-vinylporphyrin dimer 29, and 29 was found to have poor photosensitizing ability. Presumably due to the hydrophobic nature of dimers 25 and 31, these dimers were found to be almost as active as Photofrin-II@. Similar tumorcidal results were obtained in the trimer series. Moan and coworkers have also reported similar results in simple ether derivatives (with variable carbon length chains) of hematoporphyrin dicarboxylic acids.²³ Our own studies with different ether derivatives of chlorins and methyl pheophorbides have also shown that hydrophilicity/hydrophobicity characteristics play an important role for a photosensitizer to show a significant photosensitizing efficacy.²⁴ Detailed biological studies with photosensitizers discussed herein are in progress and will be reported later.

Experimental

Melting points were measured on a Thomas/Bristoline microscopic hot stage apparatus and were uncorrected. Silica gel 60 (70-230 and 230-400 mesh, Merck) or neutral alumina (Merck; usually Brockmann Grade III, i.e. deactivated with 6% water) were used for column chromatography. Preparative thin layer chromatography was canicd out on 20 x 20 cm glass plates coated with Merck G 254 silica gel (1 mm thick). Analytical thin layer chromatography was performed using Merck 60 F254 silica gel (precoated sheets, 0.2 mm thick). Reactions were monitored by thin layer chromatography and spectrophotometry and were carried out under nitrogen and in the dark. Proton and carbon-13 NMR spectra were obtained in deuteriochloroform solution at 300 MHz using a General Electric QE300 spectrometer; chemical shifts are expressed in ppm relative to chloroform (7.258 ppm). FAB mass spectra were

recorded independently at SUNY Buffalo or at Lederle Laboratories, Pearl River, New York. At Buffalo, mass spectra were obtained with an Ion-Tech fast atom gun using xenon gas on a Kratos MS80RFA instrument equipped with a Kratos DS90 data system. Compounds were dissolved in dichloromethane and 3-nitrobenzyl alcohol mixture, which served as the FAB matrix, and the solution was deposited on a stainless steel FAB probe tip. When a copper probe tip was used, protonated as well as copper-containing molecular ion species were produced. The instrument was typically operated at 1000 resolution and calibrated with sodium and cesium iodide depending on the mass range required. The samples studied at Lederle Laboratories were run on a VG ZAB-SE high performance mass spectrometer, equipped with VG11-250 data system. The instrument was calibrated with cesium iodide and scanned from m/z 350 to 2500 for low- mass samples, and from m/z 2000 to 4500 for high-mass samples. The matrix used was a 50:50:0.5 (v/v/v) thioglycerol/glycerol/TFA FAB matrix, and this was stable for the duration of the FAB experiments. Elemental analyses were obtained from Galbraith Laboratories, Knoxville,TN, USA. Electronic absorption spectra were measured in dichloromethane solution using a Hewlett-Packard 845OA spectrophotometer.

2-(l-Hydroxyethyl)-6,7-bis-(2-methoxycarbonylethyl)-l,3,S,S-tetramethylporphyrin **(11). 2-** Acetyldeuteroporphyrin-IX dimethyl ester (150 mg) was dissolved in dichloromethane (100 ml). Sodium bomhydride (380 mg) in methanol was added and the reaction mixture was stirred at room temperature for 30 min; glacial acetic acid (2 ml) was added to decompose the excess of sodium borohydride and the reaction mixture was poured into water, extracted with dichloromethane, washed with aqueous sodium bicarbonate and again with water, and then dried over anhydrous sodium sulfate. Evaporation of the solvent gave a residue which was chromatographed on a silica gel column eluted with 2% methanol in dichloromethane. The appropriate eluates were collected and evaporation of the solvent, followed by crystallization from dichloromethane/hexane gave the title compound in 90% yield (135 mg). UV-Vis λ max, 400 nm (e 169,100), 498 (13,500), 531 (8,170), 566 (6,100), 620 (3,800). NMR (ppm): 10.20, 10.00, 9.98, 9.96 (s, 1H, 4 meso-H), 9.02 (s, 1H, β -H), 6.25 [q, 1H, C<u>H</u>(OH)CH₃], 4.40 (m, 4H, 2CH₂CH₂CO₂), 3.72 (s, 6H, 2 OCH₃), 3.70, 3.68, 3.64, 3.60 (s, 3H, 4 CH₃), 3.26 (t, 4H, CH₂CH₂CO₂), 2.10 [d, 3H, CH(OH)CH₃, -4.0 (s, 2H, 2NH). Analysis Calcd for C₃₄H₃₈N₄O₅: C, 70.09; H, 6.49; N, 9.43. Found: C, 69.91; H, 6.45; N, 9.39.

4-(l-Hydroxyethyl)-6,7-bis(2-methoxycarbonylethyl)-l,3,S,8-tetramethylporphyrin (10). 4-Acetyldeuteroporphyrin-IX dimethyl ester (150 mg) was treated with sodium borohydride (380 mg) by following the procedure as described for the foregoing porphyrin and was isolated in 85% yield (128 mg), m.p. 216-218'C. UV-Vis hmax, 400 nm (e 173,400), 498 (14,000) 531 (8,500), 566 (6,300), 620 (3,900). NMR (ppm): 10.25, 10.00, 9.98, 9.90 (s, 1H, 4 meso-H), 9.05 (s, 1H, β-H), 6.25 [q, 1H, C<u>H(</u>OH)CH₃], 4.40 (m, 4H, C<u>H2</u>CH₂CO₂), 3.75 (s, 6H, 2, OCH3), 3.70, 3.52, 3.50, 3.45 (s, 3H, 4 CH3), 3.30 (t, 4H, CH2CH2CO2), 2.10 [d, 3H, CH(OH)CH3], -4.0 (s, 2H, 2 NH). Analysis Calcd for C34H3aN405: C, 70.09; H, 6.49; N, 9.43. Found: C, 69.97; H, 6.54; N, 9.49.

Porphyrin Dimer (31). 2-(1-Hydroxyethyl)-6,7-bis-(2 methoxycarbonylethyl)-1,3,5,8-tetramethylporphyrin 11 **(25** mg) was stirred with 30 % HBr/acetic acid in a stoppered flask for 2 h at room temperature. The solvent was evaporated under high vacuum (1 mm Hg). The resulting (1-bromoethyl) derivative was not isolated and was immediately treated with porphyrin **11** (25 mg) dissolved in dichloromethane (10 ml). The reaction mixture was stirred at room temperature for 10 min under a nitrogen atmosphere, diluted with dichloromethane, and then poured into cold water (200 ml). The dichloromethane layer was washed with water (4x100 ml), dilute aqueous sodium bicarbonate solution, and then with water again. The organic layer was dried over anhydrous sodium sulfate and evaporation of the solvent gave a residue which was purified on preparative silica gel thick layer plates (developed with 5% methanol in dichloromethane). Three main bands were isolated. The most labile band was identified as 2-vinyl deuteroporphyrin-IX dimethyl ester (m.p. 211-212°C), most likely formed due to dehydration of the starting porphyrin. The least polar band was identified as the starting porphyrin **11.** The central band was characterized as the desired dimer 31. The product was isolated by washing the appropriate silica gel band with 5% methanol in dichloromethane, which was then washed with water before drying over anhydrous sodium sulfate. Evaporation of the solvent and crystallization with dichloromethane/hexane gave the title dimer in 58% yield (28.5 mg). The yield was >75% if calculated on the basis of the consumed starting porphyrin. M.p. 108-112 °C. UV-Vis λ max: 398 nm (ϵ) 218,600), 497 (24,350), 529 (13,750), 568 (10,520) and 622 (7,100). NMR: (ppm), 10.40, 10.05 (s, lH, 2 meso-H); 9.98, 9.65, 9.60 (s, 2H, 6 meso-H); 8.40 (s, 2H, β-pyrrolic H), 6.70 [m, 2H, 2 CH(CH3)-O-]; 4.45 (m, 8H, 4 $CH_2CH_2CO_2$; 3.70, 3.68 (s, 6H, 4-OCH₃); 3.60 (s, 6H, CH₃); 3.39 (s, 6H, 2 CH₃); 3.30 (m, 8H, CH₂CH₂CO₂); 2.53 (s, 6H, 2 CH₃); 2.51 [d, 6H, 2 CH(OH)CH₃]; -4.35 (s, 4H, 4 NH). HRMS Calcd for C₆₈H₇₄N₈O₉; 1147. 5660 (M+l). Found: 1147.5682.

Porphyrin Dimer (25). 4-(1-Hydroxyethyl)-6,7-bis-(2-methoxycarbonylethyl)-1,3,5,8-tetramethylporphyrin 10 (25 mg) was converted into the corresponding (I-bromoethyl) derivative and then condensed with porphyrin 10 following the method described for the foregoing porphyrin. The title compound was isolated in 48% yield. M.p. 100-102⁸C. UV-Vis λ max: 400 nm (e 190,000), 498 (20,500), 528 (13,000), 565 (10,000) and 622 (6,500). NMR (ppm): 10.10, 9.75 (s, 3H, 6-meso H); 9.52 (s, 2H, 2 meso-H); 8.98 (s, 2H, p-pyrrolic H); 6.70 [q, 2H, 2 $\text{CH}(\text{CH}_3)$ -O-1; 4.40 (m, 8H, CH₂CH₂CO₂); 3.80, 3.40 (s, 3H, 2 CH₃); 3.72, 3.45 (s, 9H, 3 CH₃); 3.70 (s, 12H, 4-

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OCH₃); 3.32 (m, 8H, CH₂CH₂CO₂); 2.50 [d, 6H, 2 CH(OH)CH₃]; -4.40 (s, 4H, 4 NH). HRMS Calcd for $C_{68}H_{74}N_8O_9$; 1147.5660 (M+1). Found: 1147.5678.

Diacetylporphyrin Dimer (26).

Merhod 1: 2-Acetyl-4-(l-hydroxyethyl)-deuteroporphyrin-IX dimethyl ester 12 (25 mg) was dissolved in dry dichloromethane (10 ml). The reaction mixture was stirred at -7O'C under a nitrogen atmosphere. Freshly distilled methane sulfonyl chloride (100 μ) was added and the mixture was stirred for 2 h. Lithium bromide (20 mg) dissolved in dry THF **(5** ml) was added to the reaction mixture and it was stirred for a further 1 h. Porphyrin 12 (25 mg) dissolved in dry dichloromethane (15 ml) was added to the reaction mixture, which was stirred at room temperature for an additional 15 min. It was then diluted with dichloromethane (100 ml), washed with water, aqueous sodium bicarbonate, and again with water. Evaporation of the solvent gave a residue which was found to be a mixture of three components by analytical TLC. The most labile fraction was identified as 2-acetyl-4-vinyl-deuteroporphyr dimethyl ester. The second fraction was identified as the diacetyl dimer 26 and the most polar band was found to be the starting material 12. The diacetyl dimer was crystallized from dichloromethane/hexane and was obtained in 25% yield (13 mg) yield. M.p. 222-224°C. UV-Vis Amax: 406 nm **(E** 184,900), 510 (14,700), 548 (14,100), 578 (10,600), 636 (3,100). NMR (ppm): 10.38, 9.92, 9.08 (s, 2H, 6 meso-H), 9.58 (bs, 2H, 2 meso-H), 6.60 [q, 2H, 2C<u>H</u>(CH₃)-O-], 4.32 (m, 8H, 4 C<u>H</u>₂CH₂CO₂), 3.90, 3.75, 3.72, 3.60, 3.58, 3.50 (s, 36 H, 8 ring CH₃ and 4 OCH3), 3.20 (s, 6H, 2 COCH3), 3.18 (m, 8H, 4 CH2C<u>H2</u>CO₂), 2.58 [d, 6H, 2 CH(C<u>H</u>3)-O-], -4.92 (s, 4H, 4 NH). HRMS Calcd for C₇₂H₇₈N₈O₁₁, 1231.5868 (M+I). Found: 1231.5854.

Method 2: Porphyrin 12 (12 mg) was dissolved in dry dichloromethane (10 ml) and HBr gas was slowly bubbled through the solution for 5 min; the reaction mixture was then stirred for 15 min. The solvent was evaporated under high vacuum at 30°C. The residue was redissolved in dichloromethane (10 ml) and porphyrin 12 (12 mg) dissolved in dichloromethane (10 ml) was added and the reaction mixture was stirred at room temperature for 10 min. The mixture was diluted with dichloromethane (100 ml), washed with water, aqueous sodium bicarbonate and again with water. Evaporation of the solvent gave a residue and the title compound was purified by preparative TLC as described above, and was obtained in 50% yield (12 mg). The dimer was identical in every respect with the diacetyl dimer 12 obtained by Method 1. Method 2 was more convenient than the mesylate method, so all other dimers and trimers were prepared by following this approach.

Diacetylporphyrin Dimer (32). The (1-bromoethyl) derivative 20 prepared from corresponding (1-hydroxyethyl) analogue 13 (25 mg), was condensed with the starting porphyrin 13 (25 mg), and dimer 32 was obtained in 48% yield (24 mg). M.p.146-147'C. UV-Vis hmax: 406 nm **(E** 218,000), 512 (18,700), 548 (17,000). 636 (3,350). NMR (ppm): 10.28, 9.92, 9.64 (s, 2H, 6 meso-H), 9.50 (bs, 2H, 2 meso-H), 6.65 [q, 2H, 2 C<u>H</u>(CH₃)-O-], 4.48, 4.31 (t, 4H, CH₂CH₂CO₂), 3.75, 3.67, 3.50 (s, total 36 H, 8 CH₃ and 4 OCH₃), 3.30 (s, 6H, 2 COCH₃), 3.25 (m, 4H, CH₂CH₂CO₂), 2.55 [d, 6H, CH(CH₃)-O-], -4.07 (s, 4H, 4NH). HRMS Calcd for C₇₂H₇₈N₈O₁₁; 1231.5868 (M+l). Found: 1231.5858.

Diacetylporphyrin Dimer (36). This dimer was prepared by condensing 2-acetyl-4-(l-bromoethyl) deuteroporphyrin-IX dimethyl ester 19, obtained from 4-(1-hydroxyethyl) analogue 12 (20 mg) with 2-(lhydroxyethyl)-4-acetyldeuteroporphyrin-IX dimethyl ester 13 (20 mg) and was isolated in 52% yield (26 mg). M.p. 180-182°C. UV-Vis λ max: 406 nm (e 218,000), 510 (21,200), 548 (19,250), 580 (14,600), 636 (5,300). NMR (ppm): 10.38, 10.18, 9.90, 9.80, 9.60, 9.10 (s, lH, 6 meso-H), 9.50, 9.35 (bs, lH, 2 meso-H), 6.65 and 6.60 [q (merged), 2H, 2 CH(CH3)-O-), 4.50, 4.38 (t, 4H, 4 CH₂CH₂CO₂), 9.82, 9.78, 9.77, 9.76, 9.73, 9.72, 9.70, 9.55, 9.52 (s, total 36H, $\overline{8}$ CH₃ and 4 OCH₃), 2.55, 2.45 [d, 3H, 2 CH(CH₃)-O], -2.70 and -2.71 [s (merged), 2H, 4 NH]. HRMS calculated for $C_{72}H_{78}N_8O_{11}$; 1231.5868 (M+1). Found: 1231.5815.

Mono-acetyl-mono-vinylporphyrin Dimer (28). 2-Acetyl-4-(1-hydroxyethyl)-deuteroporphyrin-IX dimethyl ester 12 (22 mg) was converted to (1-bromoethyl) analogue 19 and condensed with 4-(l-hydroxyethyl)-2 vinyldeuteroporphyrin-IX dimethyl ester 18^{25} (22 mg) and the title dimer was obtained in 50% yield (22 mg). M.p. 212-214'C. UV-Vis hmax: 402 **nm (E** 200,9OO), 506 (14,400), 540 (1 l,OOO), 576 (8,700), 628 (2,200). NMR (ppm): 10.32, 9.99, 9.92, 9.58, 9.20, 9.17 (s, 1H, 6 meso-H), 9.80, 9.50 (bs, 1H, 2 meso-H), 8.00 (m, 1H, CH=CH₂ 6.62 [m, 2H, 2 CH(CH₃)-O-], 6.14 and 6.35 (d, 2H, CH=CH₂), 4.37 (m, 8H, 4 CH₂CH₂CO₂), 3.90, 3.76, 3.70, 3.67, 3.65, 3.55, 3.47, 3.37 (s, total 36 H, 8 CH3 and 4 OCH3), 3.21(s, 3H, COCH3), 3.20 [m (merged with acetyl protons), 8H, 4 CH₂CH₂CO₂], 2.58 [m (2 d merged), 6H, 2 CH(CH₃)-O-], -4.80 and -5.00 (s, 2H, 4 NH); HRMS Calcd for $C_{72}H_{78}N_8O_{10}$; 1215.5919 (M+1). Found:1215.5902.

Mono-acetyl-mono-vinylporphyrin Dimer (34). 4-Acetyl-2-(1-bromoethyl)-deuteroporphyrin-IX dimethyl ester 20 obtained from 13 (15 mg) was treated with 2-(1-hydroxyethyl)-4-vinyldeuteroporphyrin-IX dimethyl ester 17 25 (15 mg) and the title dimer 34 was obtained in 33% yield (10 mg). M.p.198-200°C. UV-Vis Xmax: 402 nm **(E** 208,000), 506 (15,100), 540 (ll,OOO), 578 (9,100), 626 (4,000). NMR (ppm): 10.34, 10.24, 10.08, 9.92, 9.86, 9.85, 9.63, 9.56, 9.49 (s, total 8H, 8 meso-H), 8.29-8.19 (m, 1H, CH=CH2), 6.66 [m, 1H, CH(CH3)-O-], 6.386.19 [m, 3H, CH=CH₂ and CH(CH₃)-O-], 4.47-4.30 (m, 8H, 4 CH₂CH₂CO₂), 3.73-3.46 [multiple s, 36 H, 8 CH₃ and 4 OCH3), 3.40-3.20 (m, 8H, CH₂CH₂CO₂), 2.54, 2.49 (1d and 1s merged, 6H, COCH₃ and CH(CH₃)-O-1, 2.11 [d, 3H, CH(CH₃)-O-], -4.13, -4.15, -4.21 and -4.36 (s, 1H, 4 NH). HRMS Calcd for C₇₂H₇₈N₈O₁₁; 1215.5916 (M+l). Found: 1215.5902.

Mono-acetyl-mono-vinylporphyrin Dimer (38). 2-Acetyl-4-(1-bromoethyl)-deuteroporphyrin-IX dimethyl ester 19 prepared from 12 (15 mg) upon condensation with 2-(1-hydroxyethyl)-4-vinyldeuteroporphyrin-IX dimethyl ester 17 (15 mg) gave dimer 38 in 23 % yield (7 mg). M.p. 170-172'C. UV-Vis hmax: 402 nm (E **255,000), 504 (22.7501, 538 (17,400),626 (6,700).** NMR (ppm): 10.27, 10.18, 10.05, 9.86, 9.84, 9.68, 9.50, 9.43, 9.24 (s, total 8 H, 8 meso-H), 8.24 (m, 1H, CH=CH₂), 6.69 [m, 1H, C<u>H</u>(CH₃)-O-], 6.37-6.14 [m, 3H, CH=CH₂ and CH(CH₃)- $O-I$, 4.38-4.29 (m, 8H, 4 $CH_2CH_2CO_2$), 3.73-3.38 (multiple s, 36 H, 8 CH₃ and 4 OCH₃), 3.25 (m, 8H, 4 $CH_2CH_2CO_2$), 3.18 (s, 3H, COCH₃), 2.58, 2.38, 2.08 (m, m, d [total 6 H, 2 CH(CH₃)-O-1, -4.08 (s, 2H, 2 NH), -4.65 and -4.80 (s, 1H, 2NH). HRMS Calcd for $C_{72}H_{78}N_8O_{11}$; 1215.5916 (M+1). Found: 1215.5881.

Mono-acetyl-mono-vinylporphyrin Dimer (40). 4-Acetyl-2-(1-hydroxyethyl)-deuteroporphyrin-IX dimethyl ester 13 was converted into its (1-bromoethyl) analogue 20 and was condensed with 4-(1-hydroxyethyl)-2 vinyldeuteroporphyrin-IX dimethyl ester 18 (15 mg) and dimer 40 was obtained in 40 % yield (12 mg). M.p. 205- 207°C. UV-Vis Xmax: 402 nm (E **245,000), 504 (21,100), 538 (16,200). 574 (12,800), 626 (5,800).** NMR (ppm): 10.32, 10.00, 9.90, 9.72 (s lH, 4 meso-H), 9.70,(s, 2H, 2 meso-H), 9.32 (bs, 2H, 2 meso H), 7.95 (m, lH, CH=CH₂), 6.65 [m, 2H, 2 CH(CH₃)-O-], 6.10-6.22 (m, 2H, CH=CH₂); 4.50, 4.30 (t, 4H, 4 CH₂CH₂CO₂), 3.75, 3.74, 3.72, 3.71, 3.62, 3.58, (s, total 36 H, 8 CH₃ and 4 OCH₃), 3.26 (m, 8H, 4 CH₂CH₂CO₂); 3.20 (s, 3H, COCH₃), 2.60, 2.40 [d, 3H, 2CH(C<u>H</u>₃)-O-], -4.30 and -4.40 (s, 2H, 4NH). HRMS Calcd for C₇₂H₇₈N₈O₁₁ 1215.5919 (M+l). Found: 1215.5946.

Hematoporphyrin Dimer (27). Diacetyl dimer 26 (25 mg) was dissolved in dichloromethane (20 ml). An ice chilled solution of methanol (10 ml) containing sodium borohydride (50 mg) was added and the reaction mixture was **stirred** at >5'C until completion of the reaction was determined by analytical TLC and spectrophotometry (etio- to rhodo- and back to etio-type). The reaction mixture was cooled to 0° C and dilute aqueous acetic acid was added to quench any excess of sodium borohydride. The mixture was extracted with dichloromethane, washed with water (3 x 200 ml), then dried over anhydrous sodium sulfate, and the solvent was removed under high vacuum at 3O'C. The residue was purified by preparative TLC on silica gel, eluting with 5% methanol in dichloromethane. The major band was collected. Evaporation of the solvent and crystallization with dichloromethane/hexane afforded the tide porphyrin dimer in 80% yield (20 mg). M.p. 235-237'C. UV-Vis kmax: 394 nm **(E** 200,800), 504 (20,000), 534 (12,400). 470 (11,400), 622 (5,200). NMR (ppm): 9.97, 9.95, 9.92, 9.80, 9.48, 9.38 (all s), 9.70, 9.20 [(both b) (total 8 H, 8 meso-H)], 6.65, 6.58, 6.50, 6.10, 5.50, 5.70 [bm, total 4H, 2 CH(OH)CH₃ and 2 CH(CH3)-O-1, 4.43 (m, 8H, 4 CH₂CH₂CO₂), 3.75-3.52 (10 s, 36H, 8 ring CH₃ and 4 OCH₃), 3.30, 3.10 (m, total 8H, 4 CH₂CH₂CO₂), 2.58 [m, 6H, CH(CH₃)-O-], 1.96 [m, 6H, CH(OH)CH₃], -4.76 (s, 1H, NH), -4.83 (s, 2H, NH), -4.92 (s, 1H, NH). HRMS Calcd for $C_{72}H_{82}N_8O_{11}$; 1235.6181 (M+1). Found: 1235.6152.

The other diacetyl- and mono-acetyl-dimers were reduced to the corresponding (1-hydroxyethyl) dimers upon treatment with sodium borohydride and were obtained in excellent yield. Their spectral details (UV-Vis and NMR) and physical constants are as follows:

Hematoporphyrin Dimer (33). This dimer was obtained from diacetyl dimer 32 in 82% yield. M.p.185-187°C. UV-Vis Imax: 396 nm **(E** 210,000), 504 (20,800), 534 (13,300), 570 (12,100), 622 (5,200). NMR (ppm): 10.28, 10.22, 10.10, 10.00, 9.32 (all bs), 10.22, 10.20, 10.18, 10.13 [(s) (total 8H, meso-H)], 6.60 [m, 2H, CH(OH)CH₃], 6.40 [m, 2H, CH(CH₃)-O-], 4.58 (m, 8H, 4 CH₂CH₂CO₂), 3.80-3.52(s merged, 36 H, 8 CH₃ and 4 OCH₃), 3.50 (m, 8H, CH₂CH₂CO₂), 2.50 [m, 6H, 2 CH(CH₃)-O-], 2.35 (m, 6H, CH(OH)CH₃), -4.02, -4.50 (s, 2H, 4 NH). HRMS Calcd for $C_{72}H_{82}N_8O_{11}$; 1235.6181 (M+1). Found: 1235.6193.

Hematoporphyrin Dimer (37). Diacetyl dimer 36 (25 mg) was reduced following similar approach and the tide dimer was obtained in 80% yield. M.p. 180-182°C. UV-Vis Amax: 406 nm (ε 218,000), 510 (21,200), 548 (14,700), 626 (5,300). NMR: 10.04-8.70 (multiple s, 8H, 8 meso-H), 6.32-6.10 [m, 4H, 2 CH(OH)CH3 and 2 CH(CH3)-O-], 4.50 (m, 8H, 4 C $H_2CH_2CO_2$), 3.80, 3.78, 3.73, 3.64, 3.62, 3.60, 3.58, 3.56, 3.42 (s, total 36 H, 8 CH₃ and 4 OCH₃), 3.25 (m, $\overline{8}H$, 4 CH₂CH₂CO₂), 2.52 [m, 6H, CH(CH₃)-O-], 0.90 [m, 6H, CH(OH)CH₃], -4.70, -4.82, -4.90, -5.00 (s, 1H, 4 NH). HRMS Calcd for C₇₂H₈₂N₈O₁₁; 1235.6181 (M+1). Found: 1235.6155.

Mono-(l-hydroxyethyl)-mono-vinyl Dimer (29). Mono-acetyl-mono-vinyl dimer 28 (10 mg) was treated with sodium borohydride (20 mg) in methanol (10 ml) and the title dimer 29 was afforded in 80% yield. M.p.135- 137°C. UV-Vis λ max: 404 nm (ε 257,000), 502 (22,550), 572 (16,800), 624 (9,300). NMR (ppm): 10.25-9.20 (multiple s, 8H, 8 meso-H), 8.36-8.22 (m, 1H, CH=CH2), 6.68-5.60 [m, 4H, 2CH(CH3)-O- and CH=CH2], 4.50-4.30 (m, 8H, CH₂CH₂CO₂), 3.78-3.40 (multiple s, 36 H, 8 CH₃ and 4 OCH₃), 3.22-3.25 (m, 8H,

 $CH_2CH_2CO_2$),2.60 [m, 6H, 2CH(CH₃)-O-], -3.88 (s, 2H, 2NH), -4.62 and -4.70 (s, 1H, 2NH). HRMS Calcd for $C_{72}H_{80}N_8O_{10}$; 1217.6075 (M+1). Found: 1217.6002.

Mono-(l-hydroxyethyl)-mono-vinyl Dimer (35). This dimer was obtained in 82% yield from the corresponding acetyl analogue 34 (20 mg) M.p. 127-129OC. UV-Vis Xmax: 404 nm **(E** 239,000), 504 (17,6CQ, 538 (13,200), 574 (10,000), 626 (3000). NMR (ppm): 10.25-9.45 (multiple s, 8H, 8 meso-H), 8.25 (m, 1H, CH=CH2), 6.63-6.50 [m, 2H,C<u>H</u>(OH)CH₃], 6.40-6.17 (m, 2H, CH=C<u>H2</u>), 4.52-4.25 (m, 8H, 4 C<u>H2</u>CH2CO₂), 3.79, 3.66, 3.65, 3.64, 3.63, 3.62, 3.61, 3.60, 3.58, 3.53, 3.50 (s, total 36 H, 8 CH3 and 4 OCH3), 3.40-3.20 (m, 8H, $CH_2CH_2CO_2$, 2.58, 2.43 [m, 3H, 2 CH(CH₃) -O-], 2.12 [d, 3H, CH(OH)CH₃], -4.15 (s, 2H, 2 NH), -4.42 and -4.58 (bs, 1H, 2 NH). HRMS Calcd for $C_{72}H_{80}N_8O_{11}$; 1217.6075 (M+1). Found: 1217.6068.

Mono-(l-hydroxyethyl)-mono-vinyl Dimer (39). The acetyl group of dimer 38 (30 mg) was reduced to the corresponding (I-hydroxyethyl) and the title dimer was produced in 84% yield (25 mg). M.p.134-138°C. UV-Vis λmax: 402 nm (ε 240,000), 504 (20,000), 536 (16,500), 572 (12,000), 624 (7,500). NMR (ppm): 10.32, 10.22, 9.94, 9.87, 9.85, 9.60, 9.40, 9.12 (s, total 8H, 8 meso-H), 8.28 (m, 1H, CH=CH2), 6.30-5.62 [m, 4H 2] CH(OH)CH₃ and CH=CH₂), 4.40 (m, 8H, 4 CH₂CH₂CO₂), 3.75, 3.70, 3.65 3.60, 3.57, 3.50, 3.44 (s, total 36 H, 8 CH3 and 4 OCH3), 3.30-3.20 (m, 8H, CH2CH2CO2), 2.57 [m, 6H, 2 CH(CH3)-O-], 2.18 [d, 3H, CH(OH)CH3] -3.85 (s, 2H, 2 NH), -3.70 and -3.85 (s, 1H, 2 NH). HRMS Calcd for $C_{72}H_{80}N_8O_{11}$; 1217.6075 (M+1). Found: 1217.6086.

Mono-(1-hydroxyethyl)-mono-vinyl Dimer (41). Dimer 40 (15 mg) was converted into the corresponding (1-hydroxyethyl) derivative upon treatment with sodium borohydride, and the title product was obtained in 85% yield. M.p.128-131°C. UV-Vis λ max: 406 nm (e 238,500), 504 (21,000), 536 (17,500), 570 (13,000), 626 (8,000). NMR (ppm): 10.35-9.15 (multiple s, total 8 H, 8 meso-H), 8.26 (m, 1H, C<u>H</u>=CH2), 6.32-6.12 [m, 4H, 2 C<u>H</u>(OH)CH₃ and CH=CH₂], 4.35-4.32 (m, 8H, 4 CH₂CH₂CO₂), 3.75-3.38 (multiple s, 36 H, 8 CH₃ and 4 OCH₃), 3.28-3.20 (m, 8H, 4 $\overline{\text{CH}}_2\text{CH}_2\text{CO}_2$), 2.20 [d, $3\overline{\text{H}}$, $\overline{\text{CH}}(\overline{\text{OH}})\text{CH}_3$], -3.85 to -4.60 (3 bs, 4 H, 4 NH). HRMS Calcd for $C_{72}H_{80}N_8O_{11}$; 1217.6075 (M+1). Found: 1217.6074.

Diacetylporphyrin Trimer (42). Hematoporphyrin-IX dimethyl ester (20 mg) in dichloromethane (15 ml) was treated with HBr gas as described for the synthesis of the hematoporphyrin dimer 27. The 2,4-bis-(1-bmmoethyl) porphyrin 21 so obtained was condensed with 2-acetyl-4- $(1-hydroxyethyl)$ -deuteroporphyrin-IX dimethyl ester 12 (40 mg) and after the usual work up, the crude product was purified by preparative thick layer chromatography on silica gel and the title trimer was obtained in 31% yield (19 mg) along with starting porphyrins and protoporphyrin-IX and mon-acetyl-mono-vinyl porphyrin esters as a mixture of isomers $(m/e 1217 \text{ (M+1, 100%)})$. The dimeric mixture was not further purified. Diacetyl trimer 42 was crystallized from dichloromethane/hexane as a fine powder. M.p. 185-187'C. UV-Vis hmax: 408 **nm (E** 340,000), 506 (30,800), 546 (21,800), 576 (18,200), 624 (4,500). NMR (ppm):10.60-9.38 (12 s, 12 H, 12 meso-H), 6.80, 6.40, 6.20, 5.92 [m, 4H, 4 C<u>H</u>(CH3)-O-], 4.40-4.20 (m, 12H, 6 $CH_2CH_2CO_2$), 3.80-3.35 (multiple s, 54 H, 12 CH₃ and 6 OCH₃), 3.30 (s, 6H, 2 COCH₃), 3.25 (m, 12H, 6 $CH_2CH_2CO_2$), 2.50-2.22 [m, 12H, 4 CH(C<u>H</u>3)-O], -3.80,-3.90, -4.20, -4.38 (bs, 6 H, 6 NH). HRMS Calcd for $C_{108}H_{118}N_{12}O_{16}$; 1839.8667 (M+1). Found 1839.8859.

Diacetylporphyrin Trimer (45). Reaction of 2,4-bis-(1-bromoethyl)-deuteroporphyrin-IX dimethyl ester 21, obtained from hematoporphyrin dimethyl ester (20 mg) with 4-acetyl-2-(I-hydroxyethyl)-deuteroporphyrin-IX dimethyl ester 13 afforded dimer 45 in 32 % yield (19.2 mg). M.p.171-173'C. UV-Vis Xmax: 408 nm (e 320,000), 508 (32,200), 548 (23,100), 576 (19,000),624 (5,000). NMR (ppm): 10.55, 10.50, 10.46, 9.97, 9.95, 9.94, 9.82, 9.80, 9.75, 9.71, 9.66, 9.50 (s, 1H, 12 meso-H), 6.70 [m, 2H, 2 CH(CH3)-O], 6.22-6.12 [m, 2H, 2 CH(CH3)-O-], 4.60-4.20 (m, 12 H, 6 CH₂CH₂CO₂), 3.80-3.60 (multiple s, 54 H, 12 CH₃ and 6 OCH₃), 3.45, 3.40 (s, 3H, 2 COCH3), 3.28 (m, 12 H, $CH_2CH_2CO_2$), 2.50-2.12 [m, 12 H, 4 CH(CH3)-O-], -3.70 to -4.15 (m, 6 H, 6 NH). HRMS Calcd for C₁₀₈H₁₁₈N₁₂O₁₆; 1839.8667 (M+1). Found: 1839.8746.

Hematoporphyrin Trimer (43). Diacetylporphyrin trimer 42 (20 mg) dissolved in dichloromethane (10 ml) was treated with sodium borohydride (40 mg) following the method described above for hematoporphyrin dimer 27. The reaction product after the standard work-up was purified by preparative chromatography (silica gel plates, eluted with 5% methanol in dichloromethane). The major band was isolated and the product was crystallized from dichloromethane/hexane as a dark brown powder. Yield 15 mg (75 %). M.p. 180-183'C. UV-Vis Max: 402 nm (e 353,700), 500 (38,000), 532 (24,000), 510 (19,5OO), 622 (10,600). NMR (ppm): 10.48-9.39 (m, 12 H, 12 meso-H), 6.78-5.82 [m, 4H, C<u>H</u>(CH₃)-O-], 4.29 (bm, 12 H, 6 C<u>H2</u>CH₂CO₂),3.80-3-40 (multiple s, 54 H, 12 CH3 and 6 OCH₃), 3.23 (m, 12 H, $\overline{6}$ CH₂CH₂CO₂), 2.50-1.86 [m, 18 H, 2 CH(OH)CH₃ and 4 CH(CH₃)-O], -4.04-4.30 (bs, 6 H, 6 NH). HRMS Calcd for $C_{108}H_{122}N_{12}O_{16}$: 1843.9160 (M+1). Found: 1843.9146.

Hematoporphyrin Trimer (46). Sodium borohydride reduction of diacetyl trimer 45 (25 mg) gave the title bis- (I-hydroxethyl) trimer in 77% yield (19.2 mg). M.p 183-185'C. UV-Vis hmax: 400 nm **(E** 336,000), 502 (40,500). 510 (21,600), 622 (12,000). NMR (ppm): 10.20-9.30 (m, 12 H, 12 meso-H), 6.80-5.80 [m, 4H, 4 CH(CH₃)-O-], 4.60-4.30 (m, 12 H, 6 CH₂CH₂CO₂), 3.82-3.40 (multiple s, 54 H, 12 CH₃ and 6 OCH3), 3.40-3.20 (m. 12 H. 6) $CH_2CH_2CO_2$), 2.60-2.10 [m, 18H, 2 CH(OH)CH₃ and 4 CH(CH₃)-O], -3.90 to -4.50 (m, 6 H, 6 NH). HRMS Calcd for C₁₀₈H₁₂₂N₁₂O₁₆: 1843.9160 (M+1). Found: 1843.9232.

Divinylporphyrin Trimer (44). By following a similar approach, hematoporphyrin dimethyl ester (20 mg) was converted to 2,4-bis-(1-bromoethyl)-deuteroporphyrin-IX dimethyl ester 21 which on reaction with 4-(1 hydroxyethyl)-2-vinyldeuteroporphyrin-IX dimethyl ester 18 (40 mg) afforded the title trimer in **32% yield (19.5 mg).** M.p. 135-137°C. UV-Vis λ max: 402 nm (ε 285,000), 504 (27,500), 538 (20,000), 524 (15,250), 626 (7,400). NMR (ppm): 10.10-9.45 (multiple s, 12 H, 12 meso-H), 8.02-7.90 (m, 2H, 2 CH=CH2), 6.62-5.24 [m, 4H, 4 CH(CH3)- O_1 , CH=CH₂, 4.58-4.38 (m, 12 H, 6 CH₂CH₂CO₂), 3.75-3.20 (multiple s, 54 H, 12 CH₃ and 6 OCH₃), 3.38-3.20 (m, 12 H, 6 CH₂C<u>H₂CO₂), 2.55-2.12</u> [m, 12 H, 4 CH(C<u>H3</u>)-O-J, - 3.70 to -4.48 (bs, 6 H, 6 NH). HRMS Calcd for $C_{108}H_{118}N_{12}O_{14}$: 1807.8952 (M+1). Found: 1807.8994.

Divinylporphyrin Trimer (47). Reaction of 2,4-bis-(1-bromoethyl)-deuteroporphyrin-IX dimethyl ester 21, prepared from hematoporphyrin dimethyl ester (20 mg) with 2-(l-hydroxyethyl)-4-vinyldeuteropovhyrin-IX dimethyl ester 17 (40 mg) gave the title divinylporphyrin trimer in **30%** yield (18 mg). M.p. 139-141'C. UV-Vis aax: 404 nm **(E 282,500), 502 (28,100), 538 (21,000), 526 (15,300), 626 (8,000).** NMR (ppm): 10.12-9.45 (multiple s, total 12 H, 12 meso-H), 8.01-7.90 (m, 2H, CH=CH₂), 6.65 -5.20 [m, 8H, 4 CH(CH3)-O-1 and 2 CH=CH2J, 4.60-4.35 (m, 12 H, 6 CH₂CH₂CO₂), 3.80-3.30 (multiple s, 54 H, 12 CH₃ and 6 OCH₃), 2.56-2.25 [m, 12 H, $\overline{4}$ CH(CH₃)-O- $1, -3.70$ to -4.50 (bm, 6 H, 6 NH). HRMS Calcd for $C_{108}H_{118}N_{12}O_{14}$: 1807.8952 (M+1). Found: 1807.8970.

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